CASE REPORT Open Access



Implant therapy for a patient with osteogenesis imperfecta type I: review of literature with a case report

Shamit S. Prabhu^{1,4*}, Kevin Fortier², Michael C. May³ and Uday N. Reebye⁴

Abstract

Bone fragility and skeletal irregularities are the characteristic features of osteogenesis imperfecta (OI). Many patients with OI have weakened maxillary and mandibular bone, leading to poor oral hygiene and subsequent loss of teeth. Improvements in implant therapy have allowed for OI patients to achieve dental restoration. However, there is limited available literature on implant therapy for patients with OI. The greatest challenge in the restoration process for OI patients in an outpatient setting is ensuring primary stability and osseointegration. Improvements in synthetic grafts improve successful implant placement and prevent predisposing patients to unnecessary procedures. This report details the successful restoration process of an OI type I patient's maxillary arch in addition to a review of the currently available literature.

Keywords: Osteogenesis imperfecta type I, Implant therapy, Brittle bone disease

Introduction

Osteogenesis imperfecta (OI), colloquially known as "brittle bone disease," is a broad term for a group of congenital disorders affecting the connective tissue resulting in a susceptibility to fractures. In 1979, Sillence et al. conducted an epidemiological and genetic study of OI patients [1]. These patients were grouped according to four distinct syndromes: (1) dominantly inherited OI with blue sclerae, (2) lethal perinatal OI with radiographically crumpled femora and beaded ribs, (3) progressively deforming OI with normal sclerae, or (4) dominantly inherited OI with normal sclerae [1]. These groupings would later become the clinical features in identifying OI types I–IV. Since then, additional types of OI have been classified based on allelic heterogeneity, histological variance, radiological features, and clinical manifestations (Table 1).

In studies conducted in Europe and the US, the birth prevalence of OI was estimated to be 0.3–0.7 per 10,000 births [2, 3]. Incidence in males and females is roughly equal. The pathophysiology for OI type I is characterized

Clinically, patients with OI type I present with an increased risk of bone fractures due to fragile bone, osteoporosis, blue sclerae, short stature, joint hypermobility, and susceptibility to conductive hearing loss progressing from adolescence to adulthood [6]. OI type I can be further categorized based on the presence, Ia, or absence, Ib, of dentinogenesis imperfecta (DI) [7]. Patients with DI will have opalescent teeth due to abnormal dentin exposure through the translucent enamel with a variable blue-gray or yellow-brown hue. Radiographical features of dentinogenesis imperfecta include deposition of dentin resulting in a marked reduction of the pulp chamber and root canals, short roots with constricted

Full list of author information is available at the end of the article



by mutations in the genes for $\operatorname{pro}\alpha 1$ chains on $\operatorname{COL1A1}$ on chromosome 17 or for $\operatorname{pro}\alpha 2$ chains on $\operatorname{COL1A2}$ on chromosome 7 [4]. The prominence of type I collagen in the extracellular matrix of bones and skin results in patients with OI having qualitative or quantitative defects. In OI type I, individuals have quantitative defects in their normal type I collagen in that the collagen is functionally normal but produced in smaller quantities. Individuals with qualitative defects produce structurally defective type I collagen resulting in moderate deformations as seen in OI type IV, to severe deformation as seen in OI type III, and can even be lethal in OI type II [5].

^{*} Correspondence: shamitprabhu@gmail.com

¹Wake Forest School of Medicine, Winston-Salem, USA

⁴Triangle Implant Center, 5318 NC Highway 55, Suite 106, Durham, NC

Table 1 Osteogenesis imperfecta classifications

Type	Inheritance	Gene	Locus	Clinical features	OMIM
I [1, 4]	AD	COL1A1 or COL1A2	17q21.33 or 7q21.3	Variable bone fragility, moderate bone deformity, blue sclerae, possible dentinogenesis imperfecta	166,200
II [1, 26]	AD	COL1A1 or COL1A2	17q21.33 or 7q21.3	Perinatally lethal	166,210
III [1, 27]	AD	COL1A1 or COL1A2	17q21.33 or 7q21.3	Severe bone fragility, progressively deforming, normal sclerae, dentinogenesis imperfecta, cardiovascular complications, spinal curvature, kyphoscoliosis	259,420
IV [1]	AD	COL1A1 or COL1A2	17q21.33 or 7q21.3	Moderate bone fragility, moderate deformity, normal sclerae, short stature, possible dentinogenesis imperfecta, kyphoscoliosis	166,220
V [28, 29]	AD	IFITM5	11p15.5	Moderate to severe bone fragility, radial head dislocation, normal to blue sclerae, normal dentin	610,967
VI [30]	AR	SERPINF1	17p13.3	Moderately to severe deformity, fish-scale pattern of lamellae, excessive osteoid, normal dentin	613,982
VII [31]	AR	CRTAP	3p22.3	Severe bone fragility, progressively deforming, normal sclerae, severe rhizomelia and coxa vera, normal dentin	610,682
VIII [32]	AR	LEPRE1	1p34.2	Severe bone fragility, normal sclerae, bulbous metaphyses, round face, short barrel-shaped chest	610,915
IX [33]	AR	PPIB	15q22.31	Severe bone deformity, gray sclerae	259,440
X [34]	AR	SERPINH1	11q13.5	Multiple bone deformities and fractures, osteopenia, dentinogenesis imperfecta, blue sclerae	613,848
XI [35]	AR	FKBP10	17q21.2	Mild to severe bone deformity, normal to gray sclerae	610,968
XII [36]	AR	SP7	12q13.13	Mild bone deformity, normal dentin, normal hearing, normal sclerae	613,849
XIII [37]	AR	BMP1	8p21.3	Severe growth deficiency, severe bone deformity, normal dentin, light blue sclerae	614,856
XIV [38]	AR	TMEM38B	9q31.2	Variable bone deformity, variable osteopenia, normal dentin, normal sclerae, normal hearing	615,066
XV [39, 40]	AR	WNT1	12q13.12	Severe bone deformity, short stature, early and recurrent fractures, normal dentin, possible blue sclerae, normal hearing	615,220
XVI [41]	AR	CREB3L1	11p11.2	Severe bone deformity, beaded ribs, callus formation, cardiac irregularities	616,229
XVII [42]	AR	SPARC	5q33.1	Progressive severe bone fragility, kyphoscoliosis, mild joint hyperlaxity, short stature	616,507

AD autosomal dominant, AR autosomal recessive, OMIM Online Mendelian Inheritance in Man

corono-radicular junctions, and bulbous crowns [8, 9]. Improper dentin formation predisposes patients to an increased risk of dental fractures and increased wear on teeth, subsequently requiring corrective dental procedures [10]. Navigating treatment options for patients with OI type I pose many challenges for dental professionals. In particular, successful dental implant treatment is difficult to achieve due to requiring strong, dense bone for

acceptance of the implant. To avoid implant failure, patients must maintain routine oral care in addition to closely monitoring bone healing around the implant site. Implant treatment is even more challenging if the patient is prescribed bisphosphonates. These drugs are often administered to reduce osteoclast activity to limit bone resorption, subsequently improve bone microarchitecture, and bone density and correct vertebral size and shape



Fig. 1 Characteristic blue sclerae



Fig. 2 Pre-operative panoramic radiograph

[11–13]. One randomized, double-blind, placebo-controlled trial on the effectiveness of Risedronate in children with OI showed a significant reduction in the risk of fractures [13]. While bisphosphonate treatment may assist in the prevention of long bone fractures, it can be a detriment in the oral restoration process.

In this report, we focus on the 3-year dental implant therapy and restorative process of a 53-year-old male patient diagnosed with OI type I.

Case presentation

Evaluation

A 53-year-old male diagnosed with OI type I was referred to our clinic for extraction of the remaining maxillary teeth and evaluation for full arch immediate load hybrid prosthesis. His clinical history included osteogenesis type 1, bipolar disorder, alopecia, and hypothyroidism. The patient presented with normal stature, measuring 170.18 cm and weighing 81.65 kg with characteristic blue sclerae of OI type I (Fig. 1). Throughout his life, he has had multiple orthopedic fractures due to his OI. At the time of surgery, he was on Lamictal, Xarelto, Synthroid, lisinopril, and hydrochlorothiazide.

Extraoral, TMJ, intraoral soft tissue, and lymph node examinations produced no abnormal findings. An examination of the dentition revealed the maxillary teeth were in poor repair with a fixed bridge extending from site number 2 to site number 5 with site number 3 serving as the pontic abutment. Sites number 8, number 9, number 10, and number 11 have periodontal involvement as well as recurrent decay. He was edentulous on the posterior left maxillary arch. His lower dentition consisted of sites number 19 through number 27 with number 28 being edentulous and number 29 having a root fracture (Fig. 2). The upper jaw had good ridge width with reproducible centric relation and centric occlusion. The patient was otherwise healthy apart from medical issues directly related to his OI.

Due to his significant gag reflex, he was unable to wear a removable prosthesis. Lengthy conversations regarding implant therapy and implant options were reviewed as well as risks with his OI. Options presented included no treatment, placement of fixtures to support a removable prosthesis, placement of fixtures to support a fixed hybrid, and placement of axial implants for fixed denture prosthesis. He elected for a fixed denture prosthesis. Our

Table 2 Chronological timeline of the implant therapy of the maxilla

Date	Site number	Implant diameter (mm)	Implant length (mm)	Immediate load	Bone graft augmentation
3/26/14	12	4.3	10	Yes	Allograft
3/26/14	14	4.3	10	Yes	None
11/10/14	10	3.5	13	Yes	Allograft
3/5/15	7	3.5	13	Yes	Allograft
4/19/16	11	4.3	11.5	Yes	None
2/22/17	3	4.3	10	Yes	Allograft
2/22/17	4	5.0	10	Yes	Allograft
2/22/17	6	4.3	13	Yes	Allograft
2/22/17	8	3.5	10	Yes	Allograft
2/22/17	9	3.5	10	Yes	Allograft



Fig. 3 Post-operative panoramic radiograph

patient was apprehensive towards having full edentulation and implant placement completed all at once and decided to have the implants placed in stages (Table 2).

Surgical technique

The patient underwent implant therapy in stages under general anesthesia with immediate load protocol. Intravenous access was obtained, and the patient was anesthetized under general anesthesia by our anesthesiologist. Carpules of 2% lidocaine with 1:100,000 epinephrine, 4% articaine hydrochloride with 1:100,000 epinephrine (Septocaine), and 0.5% bupivacaine hydrochloride with 1:200,000 epinephrine (Marcaine) were used as needed. For each site, a 15 blade was used to make a sulcular incision from the mesial to the distal aspect of the tooth. A full thickness mucoperiosteal flap was elevated with a periosteal elevator exposing the buccal alveolus. Buccal bone was removed using a surgical fissure bur to allow for osteotomes and elevators to atraumatically elevate and deliver the teeth, while preserving lingual, mesial, and distal walls. Next, a straight elevator was positioned between the alveolus and the root surface. The tooth was elevated, and the periodontal ligament was separated from the alveolus. The tooth was extracted using a no. 150 upper universal forcep. The socket was curetted and irrigated with copious amounts of normal saline solution. A bone file and rongeur were used to smoothen the alveolus.

To deliver implants, all bony walls were checked with a perio probe to verify the depth. A series of osteotomy burs were used at 1000 RPM and 50 Ncm of torque with copious sterile normal saline irrigation. At each step, angulation was checked. Once the final osteotomy was completed, the site was checked to verify that all bony walls were stable. A NobelActive implant was torqued into position at greater than 30 Ncm followed by placement of a cover screw. In instances where grafting was necessary, the graft material was positioned to obliterate the bony defect using a periosteal elevator and curette to place in the bony voids. The gingival tissues were repositioned using an Adson Tissue Forcep. A tension-free closure was attained with a periosteal release technique. The sites were closed with interrupted 3-0 gut sutures. All procedures were accomplished without any further complications.



Fig. 4 Post-operative frontal view with teeth in occlusion



Fig. 5 Post-operative lateral view of the left maxillary arch

Prosthetic procedure

The standard immediate loading procedures were followed as the patient met the guidelines of a minimum torque value of 35 Ncm. All fixtures placed had intraoperative open tray impressions taken. Impressions were sent to the laboratory, and fabrication screw-retained temporary was completed. Temporaries were placed within 24 h of surgery and were torqued at 15 Ncm. Following a 6-month period of functioning in temporary prostheses, final impressions were taken via open-tray technique. He was placed in his final prostheses with no complications. Our patient settled on final prostheses consisting of a four-unit bridge cemented at sites number 3 through number 6; individual crowns placed at sites number 7, number 8, number 9, number 10, and number 11; and a screw-retained, three-unit bridge placed at sites number 12 through number 14 (Figs. 3, 4, 5, 6, and 7). The restorative dentist placed a polymethyl methacrylate (PMMA) prosthesis on the left side, and our patient will transition to his final crowns once he is financially ready.

Follow-up

Regular hygiene visits show that our OI patient has greatly improved his overall home care routine. No areas of gingival inflammation were found. Probing depths have remained 2–4 mm with no bleeding or purulent drainage at the fixtures sites. There have been no issues with implant mobility, and all healing post-operatively was uneventful.

Discussion

The vast majority of published articles regarding OI type I revolve around fractures of the long bones and treatment strategies. An extensive literature search for manuscripts detailing the implant therapy for patients diagnosed with OI produced a marginal amount of literature (Table 3). Our case posits that oral restoration is attainable without implant failure for OI type I patients. In OI type I, the collagen produced is of normal quality but in reduced quantities [14]. As a result, OI type I is considered the mildest form of OI with the majority of fractures occurring in childhood and adolescence as the



Fig. 6 Post-operative lateral view of the right maxillary arch



Fig. 7 Post-operative occlusal photograph of the maxilla

bones continue to grow. Since the collagen is of normal quality, successful osseointegration of implants can be attained with proper planning. To account for poor bone strength, Marx et al. proposed using implants as a "tent-pole" for bone graft to be placed around to consolidate and maintain the graft's volume [15].

The same factors that must be considered when placing implants in any patient are also pertinent to OI patients. However, extra emphasis should be placed on bone quantity and bone quality. In placing implants for our patient, we ensured that all fixtures attained a final torque value greater than 35 Ncm. Traditional endosseous implants require a bone healing period post-extraction of 3 months for the mandible and 6 months for the maxilla before the implant can be loaded [16]. Innovation in implant technology allows for immediate implant loading following extraction due to design changes that provide a stronger mechanical connection to the surrounding tissue [17]. While these innovations have made implant delivery much more time-effective, primary stability can be challenging in

patients with diminished bone quantity and quality. Bone graft augmentation can be utilized to ensure the osseointegration of the implant and has been utilized to achieve positive results in some OI cases [18-22]. However, some cases found successful osseointegration without the usage of bone grafts, including some of the implants placed in our patient [23, 24]. While we were able to successfully deliver implants using synthetic grafting material or no grafting material, other literature utilized autogenous bone from either the ascending ramus [21] or iliac crest [18-20, 22]. In determining the success rate of dental implants, there is a great deal of variability due in part to the varying degrees of bone quality and quantity in the OI subtypes, patient compliance to treatment plans and dental care, and a multitude of other factors typically involved in implant therapy. One retrospective and prospective study cites strong success rates in implant delivery for OI patients with a survival rate between 93 and 100% [25]. Our patient is now 4 years post-placement of his first implant procedure and has been functioning without any issues. The diagnosis of OI type I should not be a contraindication of implant therapy as our case, and others [18-25], have shown. This case differs from other cases in utilizing synthetic grafts to aid in stability and providing another case to illustrate the advancements in implant delivery for patients with bone abnormalities.

Conclusion

In conclusion, this case shows that implant therapy for patients with OI type I is a viable treatment option with appropriate planning, surgical skill, and routine care. Advancements in the fields of implants, prosthetics, and bone grafting will continue to make implants an increasingly practical treatment option for patients with OI. However, dental practitioners should always take great precaution in ensuring that bone quality and quantity is appropriate to ensure primary stability and successful osseointegration.

Table 3 Reported implant therapy for patients diagnosed with osteogenesis imperfecta

Reference	Age (years)	Gender	OI type	Bone graft augmentation	Number of implants	Implant location	Implant type
Friberg [23]	51	F	N/A**	No	6	Full maxilla	Regular platform TiUnite Brånemark System
Wannfors [18]	30	F	III	Yes	4	Full mandible	OsseoSpeed
Payne [19]	34	F	IV	Yes	11	Full maxilla and mandible	Brånemark Mk III Ti-Unite
Prabhu [24]	32	Μ	IV	No	11	Full maxilla and mandible	Brånemark titanium bone-tapped
Binger [20]	32	F	N/A**	Yes	5	Full maxilla	ITI dental standard
Lee [21]	43	F	III	Yes	2	Right posterior mandible	Paragon Screwvent internal hexed
Zola [22]	32	М	N/A**	Yes	13	Left and right posterior maxilla and left and right posterior mandible	Not specified

^{**}OI type was not specified

Abbreviation

Ol: Osteogenesis imperfecta

Acknowledgements

Not applicable

Funding

Not applicable

Availability of data and materials

Not applicable

Authors' contributions

Primary author SSP drafted the manuscript, revised the manuscript, and assisted in the surgical implant procedures. Second author KF assisted in the revision of the manuscript and tables. Third author MCM revised the manuscript and participated in drafting the literature review. Fourth author UNR carried out all surgical procedures and coordinated the report. All authors have read and approved of this manuscript.

Ethics approval and consent to participate

Not applicable

Consent for publication

Consent to publish was obtained from all applicable parties.

Competing interests

Shamit S. Prabhu, Kevin Fortier, Michael C. May, and Uday N. Reebye declare that they have no competing interests.

Publisher's Note

Springer Nature remains neutral with regard to jurisdictional claims in published maps and institutional affiliations.

Author details

¹Wake Forest School of Medicine, Winston-Salem, USA. ²Boston University Henry M. Goldman School of Dental Medicine, Boston, USA. ³Virginia Commonwealth University School of Dentistry, Richmond, USA. ⁴Triangle Implant Center, 5318 NC Highway 55, Suite 106, Durham, NC 27713, USA.

Received: 11 June 2018 Accepted: 11 October 2018 Published online: 23 November 2018

References

- Sillence DO, Senn A, Danks DM. Genetic heterogeneity in osteogenesis imperfecta. J Med Genet. 1979;16:101–16.
- Orioli IM, Castilla EE, Barbosa-Neto JG. The birth prevalence rates for the skeletal dysplasias. J Med Genet. 1986;23:328–32.
- Stevenson DA, Carey JC, Byrne JL, Srisukhumbowornchai S, Feldkamp ML. Analysis of skeletal dysplasias in the Utah population. Am J Med Genet A. 2012;158A:1046–54.
- Byers PH. Brittle bones-fragile molecules: disorders of collagen gene structure and expression. Trends Genet. 1990;6:293–300.
- Sillence DO, Rimoin DL, Danks DM. Clinical variability in osteogenesis imperfecta-variable expressivity or genetic heterogeneity. Birth Defects Orig Artic Ser. 1979;15:113–29.
- Van Dijk F, Sillence D. Osteogenesis imperfecta: clinical diagnosis, nomenclature and severity assessment. Am J Med Genet A. 2014;164:1470–81.
- Levin LS, Brady JM, Melnick M. Scanning electron microscopy of teeth in dominant osteogenesis imperfecta. Am J Med Genet. 1980;5:189–99.
- Rios D, Vieira AL, Tenuta LM, Machado MA. Osteogenesis imperfecta and dentinogenesis imperfecta: associated disorders. Quintessence Int. 2005;36:695–701.
- Bailleul-Forestier I, Berdal A, Vinchier F, de Ravel T, Fryns JP, Verloes A. The genetic basis of inherited anomalies of the teeth. Part 2: syndromes with significant dental development. Eur J Med Genet. 2008;51:383–408.
- American Academy of Pediatric Dentristy. Guideline on dental management of heritable dental developmental anomalies. Pediatr Dent. 2016;38:302–7.
- Land C, Rauch F, Montpetit K, Ruck-Gibis J, Glorieux FH. Effect of intravenous pamidronate therapy on functional abilities and level of ambulation in children with osteogenesis imperfecta. J Pediatr. 2006;148:456–60.

- Rauch F, Munns C, Land C, Glorieux FH. Pamidronate in children and adolescents with osteogenesis imperfecta: effect of treatment discontinuation. J Clin Endocrinol Metab. 2016;91:1268–74.
- Bishop N, Adami S, Ahmed SF, Anto'n J, Arundel P, Burren CP, Devogelaer J, et al. Risedronate in children with osteogenesis imperfecta: a randomised, double-blind, placebo-controlled trial. Lancet. 2013;382:1424–32.
- Marini J, Forlino A, Bächinger H, Bishop N, Byers P, Paepe A, et al. Osteogenesis imperfecta. Nat Rev Dis Primers. 2017;3:17052.
- Marx RE, Shellenberger T, Wimsatt J, Correa P. Severely resorbed mandible: predictable reconstruction with soft tissue matrix expansion (tent pole) grafts. J Oral Maxillofac Surg. 2002;60:878–88.
- Brånemark Pl. Osseointegration and its experimental background. J Prosthet Dent. 1983;50:399–410.
- Javed F, Ahmed HB, Crespi R, Romanos GE. Role of primary stability for successful osseointegration of dental implants: factors of influence and evaluation. Interv Med Appl Sci. 2013;5:162–7.
- Wanfors K, Johansson C, Donath K. Augmentation of the mandible via a "tent-pole" procedure and implant treatment in a patient with type III osteogenesis imperfecta: clinical and histologic consideration. Int J Oral Maxillofac Implants. 2009;24:1144–8.
- Payne M, Postlethwaite K, Smith D, Nohl F. Implant-supported rehabilitation of an edentate patient with osteogenesis imperfecta: a case report. Int J Oral Maxillofac Implants. 2008;23:947–52.
- Binger T, Rucker M, Spitzer WJ. Dentofacial rehabilitation by osteodistraction, augmentation and implantation despite osteogenesis imperfecta. Int J Oral Maxillofac Surg. 2006;35:559–62.
- Lee C, Ertel S. Bone graft augmentation and dental implant treatment in a
 patient with osteogenesis imperfecta: review of the literature with a case
 report. Implant Dent. 2003;12:291–3.
- 22. Zola MB. Staged sinus augmentation and implant placement in a patient with osteogenesis imperfecta. J Oral Maxillofac Surg. 2000;58:443–7.
- Friberg B. Brånemark system implants and rare disorders: a report of six cases. Int J Periodontics Restorative Dent. 2013;33:139–48.
- Prabhu N, Stevenson A, Cameron A. The placement of osseointegrated dental implants in a patient with type IV B osteogenesis imperfecta: a 9-year followup. Oral Surg Oral Med Oral Pathol Oral Radiol Endod. 2007;103:349–51.
- Jensen JL, Brox HT, Storhaug K, Ambjørnsen E, Støvne SA, Bjørnland T. Dental implants in patients with osteogenesis imperfecta: a retrospective and prospective study with review of the literature. Oral Surg. 2011;4:105–14.
- Barnes AM, Chang W, Morello R, Cabral WA, Weis M, Eyre DR, et al. Deficiency of cartilage-associated protein in recessive lethal osteogenesis imperfecta. New Eng J Med. 2006;355:2757–64.
- 27. Radunovic Z, Wekre LL, Diep LM, Steine K. Cardiovascular abnormalities in adults with osteogenesis imperfecta. Am Heart J. 2011;161:523–9.
- 28. Glorieux FH, Rauch F, Plotkin H, Ward L, Travers R, Roughley P, et al. Type V osteogenesis imperfecta: a new form of brittle bone disease. J Bone Miner. 2000;15:1650–8.
- Cho TJ, Lee KE, Lee SK, Song SJ, Kim KJ, Jeon D, et al. A single recurrent mutation in the 5-prime UTR of IFITM5 causes osteogenesis imperfecta type V. Am J Hum Genet. 2012;91:343–8.
- Glorieux FH, Ward LM, Rauch F, Lalic L, Roughley PJ, Travers R. Osteogenesis imperfecta type VI: a form of brittle bone disease with a mineralization defect. J Bone Miner. 2002;17:30–8.
- Ward LM, Rauch F, Travers R, Chabot G, Azouz EM, Lalic L, et al. Osteogenesis imperfecta type VII: an autosomal recessive form of brittle bone disease. Bone. 2002;31:12–8.
- Cabral WA, Chang W, Barnes AM, Weis M, Scott MA, Leikin S, et al. Prolyl 3hydroxylase 1 deficiency causes a recessive metabolic bone disorder resembling lethal/severe osteogenesis imperfecta. Nat Genet. 2007;39:359–65.
- Van Dijk FS, Nesbitt IM, Zwikstra EH, Nikkels PGJ, Piersma SR, Fratantoni SA, et al. PPIB mutations cause severe osteogenesis imperfecta. Am J Hum Genet. 2009;85:521–7.
- 34. Christiansen HE, Schwarze U, Pyott SM, AlSwaid A, Al Balwi M, Alrasheed S, et al. Homozygosity for a missense mutation in SERPINH1, which encodes the collagen chaperone protein HSP47, results in severe recessive osteogenesis imperfecta. Am J Hum Genet. 2010;86:389–98.
- Alanay Y, Avaygan H, Camacho N, Utine GE, Boduroglu K, Aktas D, et al. Mutations in the gene encoding the RER protein FKBP65 cause autosomalrecessive osteogenesis imperfecta. Am J Hum Genet. 2010;86:551–9.
- 36. Lapunzina P, Aglan M, Temtamy S, Caparros-Martin JA, Valencia M, Leton R, et al. Identification of a frameshift mutation in Osterix in a

- patient with recessive osteogenesis imperfecta. Am J Hum Genet. 2010; 87:110–4.
- Martinez-Glez V, Valencia M, Caparros-Martin JA, Aglan M, Temtamy S, Tenorio
 J, et al. Identification of a mutation causing deficient BMP1/mTLD proteolytic
 activity in autosomal recessive osteogenesis imperfecta. Hum Mutat. 2012;33:
 343–50.
- Shaheen R, Alazami AM, Alshammari MJ, Faqeih E, Alhashmi N, Mousa N, et al. Study of autosomal recessive osteogenesis imperfecta in Arabia reveals a novel locus defined by TMEM38B mutation. J Med Genet. 2012;49:630–5.
- Keupp K, Beleggia F, Kayserili H, Barnes AM, Steiner M, Semler O, et al. Mutations in WNT1 cause different forms of bone fragility. Am J Hum Genet. 2013;92:565–74.
- Pyott SM, Tran TT, Leistritz DF, Pepn MG, Mendelsohn NJ, Temme RT, et al. WNT1 mutations in families affected by moderately severe and progressive recessive osteogenesis imperfecta. Am J Hum Genet. 2013;92:590–7.
- 41. Symoens S, Malfait F, D'hondt S, Callewaert B, Dheedene A, Steyaert W, et al. Deficiency for the ER-stress transducer OASIS causes severe recessive osteogenesis imperfecta in humans. Orphanet J Rare Dis. 2013;8:154.
- Mendoza-Londono R, Fahiminiya S, Majewski J, Care4Rare Canada Consortium, Tetreault M, Nadaf J, et al. Recessive osteogenesis imperfecta caused by missense mutations in SPARC. Am J Hum Genet. 2015;96:979–85.

Submit your manuscript to a SpringerOpen journal and benefit from:

- ► Convenient online submission
- ► Rigorous peer review
- ► Open access: articles freely available online
- ► High visibility within the field
- ► Retaining the copyright to your article

Submit your next manuscript at ▶ springeropen.com